

REVIEW

TRITERPENOID SAPONINS AND THEIR GENINS*

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A REVIEW on triterpenoid saponins and sapogenins was published seven years ago.¹ Since then, due to improved techniques in the isolation and structural elucidation of saponins and their biological evaluation, considerable data has been accumulated on these compounds and this is covered, in the present review.

Some general reviews² on the distribution of saponins in plant kingdom and their properties have appeared during the period 1966–1972 and many new saponins have been found which possess important antimicrobial and antimycotic properties.³ Further many saponins have been isolated, which possess the same sapogenin but exhibit a very wide variation in the number, configuration and type of carbohydrate units. Oleanolic acid-containing saponins, for example, are now known to have a range of 1–10 sugar units. The period has witnessed a noteworthy progress in the isolation of pure saponins and their complete structure elucidation including the configuration of the carbohydrate moieties.

Important advances have also been registered in our understanding of the biogenetic pathways of the synthesis of saponins in plants. The role of squalene as the progenitor of polycyclic triterpenoids is now well established. The intermediacy of 2,3-epoxysqualene in lanosterol and cholesterol biogenesis in mammalian tissues and its universal intermediacy in the formation of C₃-oxygenated triterpenes has been amply substantiated. However, in the triterpenoids found in primitive plants, which have no oxygen function at C₃, it has been shown that their biogenesis may involve a direct proton-initiated cyclization of squalene.⁴

DETECTION AND ISOLATION

The techniques reported in the earlier review,¹ have continued to be employed for the detection and isolation of saponins and no significant advance has been made in this area

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¹ BASU, N. and RASTOGI, R. P. (1967) *Phytochemistry* **6**, 1249.

² HILLER, K., KEIPERT, M. and LINZER, B. (1966) *Pharmazie* **21**, 713; WULF, G. (1968) *Deut. Apoth.-Ztg.* **108**, 797; WOITKE, H. O., KAYSER, J. P. and HILLER, K. (1970) *Pharmazie* **25**, 212.

³ PLERICIC, J. (1970) *Bilt. Farm. Drus. Soc. Repub. Makedonija*, **16**, 199; CUCU, V. and GRECU, L. (1971) *Farma-cia* **19**, 641, (1972) *Chem. Abstr.* **76**, 81591; CHEFFE, P. R. (1971) *Can. J. Animal Sci.* **51**, 621.

⁴ COREY, E. J. and ORITZ DE MONTELLANO, P. R. (1967) *J. Am. Chem. Soc.* **89**, 3362; GODFREDSON, W. O., LORK, H., VAN TAMELEN, E. E., WILLET, J. D. and CLAYTON, R. B. (1968) *ibid.*, **90**, 208; EPPENBURGER, U., HIRTH, L. and OURISSON, G. (1969) *European J. Biochem.* **8**, 180; PONSINET, G. and OURISSON, G. (1968) *Phytochemistry* **7**, 757; BARTON, D. H. R., MELLOWS, G. and WIDDONSON, D. A. (1971) *J. Chem. Soc. (C)* 1110.

during the period under review. Saponin mixtures have been separated by two dimensional chromatography on silica gel H plates⁵ and periodate-alkaline potassium permanganate (Lemeux reagent)⁶ and antimony trichloride-vanillin (5:4) in chloroform have been used for their detection. The antimony pentachloride procedure for the colorimetric estimation of saponins has been studied to determine the optimal conditions for the development of colour.⁷ The saponin content of the plants belonging to the Chenopodiaceae has been estimated by the hemolytic index method.⁸ The distribution of saponins in *Bupleurum falcatum* roots has been visualized histochemically with 99% EtOH-conc. H_2SO_4 (1:1) or Fe_2SO_4 .⁹

The acidic saponins of *Sapindus mukorossi* have been successfully precipitated as their ammonium salts¹⁰ from crude extracts with $(\text{NH}_4)_2\text{SO}_4$. For purification, saponins have been separated on silica gel¹¹ using the solvent system BuOH-EtOAc- H_2O (1:1:1) and alumina¹² with BuOH-EtOAc-25% NH_4OH (7:2:5).¹³

CHEMICAL STRUCTURE

The aglycone fraction of hydrolysed saponins has been very often found to be a mixture of genins which are produced during drastic conditions of acid hydrolysis. Mild acid hydrolysis of glycosides from *Aegiceras corniculatum* by the Barton procedure, therefore, has been carried out to obtain an improved yield of the genuine sapogenin.¹⁴ By varying hydrolytic procedures, artefact and genuine sapogenins have been obtained as in the cases of aescigenin¹⁵ and protoaescigenin¹⁶ from *Aesculus hippocastanum*, cyclamiretins D & A from *Cyclamen* sp.,¹⁷ panaxadiol¹⁸ and protopanaxadiol¹⁹ from the *Ginseng* roots and senegenin²⁰ (or polygallic acid = senegenic acid)²¹ and presenegenin²² from the *Senega* roots. The genuine sapogenins have been obtained either under mild acidic conditions,^{16,17} by chemical degradative procedures such as halogenation in case of protopanaxadiol,¹⁹ periodate oxidation followed by alkaline cleavage in cases of presenegenin²² and saikogenins²³. A hydrolytic enzyme(s) isolated in a partially purified state from the

⁵ KARTNIG, TH. and WEGSCHÄIDER, O. (1972) *Planta Med.* **21**, 144; KARTNIG, TH., WEGSCHÄIDER, O. and BL. C. Y. (1972) *ibid.* **21**, 29.

⁶ RAMCHANDRA ROW, L. and RUKMINI, C. (1966) *Indian J. Chem.* **4**, 149.

⁷ KISHIHARA, S. and SIMIZU, T. (1967) *Kobe Daigaku Nogakubu Kenkyu Hokoku* **8**, 31; (1969) *Chem. Abstr.* **70**, 39080 w.

⁸ MELKILMYAN, K. H. A., BEVAZOVA, L. V. and SERBYAN, S. E. (1969) *Zh. Akad. Nauk.* **22**, 94; (1970) *Chem. Abstr.* **72**, 28844 p.

⁹ NAGOSHI, K., ODANI, T. and HIGASHI, J. (1970) *Shoyakuquaku Zasshi* **24**, 93; (1971) *Chem. Abstr.* **75**, 72521.

¹⁰ RAMACHANDRA ROW, L. and RUKMINI, C. (1966) *Indian J. Chem.* **4**, 36.

¹¹ ISMAILOV, A. I. and ISKENDROV, G. B. (1969) *Akadem. Biol. Nauka* **11**, 95; (1970) *Chem. Abstr.* **72**, 63584 a.

¹² BUKHAROV, V. G. and KARNELVA, L. N. (1970) *Izv. Akad. Nauk. SSSR, Ser. Khim.* **171**; (1970) *Chem. Abstr.* **73**, 4146 g.

¹³ MUKHAMEDZIEV, M. M. and ALIMBAEVA, P. K. (1970) *Farmatsiya* **19**, 39; (1970) *Chem. Abstr.* **73**, 117152 z.

¹⁴ HENSENS, O. D. and LEWIS, K. G. (1965) *Tetrahedron Letters* **4639**.

¹⁵ CAINELLI, G., MELERA, A., ARIGONI, D. and JEGGER, O. (1957) *Helv. Chim. Acta* **40**, 2390.

¹⁶ KUHN, R. and LOEW, I. (1963) *Liebigs Ann.* **669**, 183.

¹⁷ TSCHESCHE, R., INCHANNRONDONO, F. and WULFF, G. (1964) *Liebigs Ann.* **680**, 107; TSCHESCHE, R., STRIEGLER, H. and FRIHLHABER, H. W. (1966) *ibid.* **691**, 165.

¹⁸ TANAKA, O., NAGAI, M. and SHIBATA, S. (1964) *Tetrahedron Letters* **2291**.

¹⁹ SHIBATA, S., TANAKA, O., ANDO, T., SADO, M., TSUSHIMA, S. and OHSAWA, T. (1966) *Chem. Pharm. Bull.* **14**, 595.

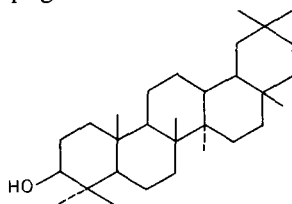
²⁰ DUGAN, J. J., DE MAYO, P. and STARRATT, A. N. (1964) *Can. J. Chem.* **42**, 491.

²¹ PELLTIER, S. W., ADITYACHAUDHURY, N., TOMASZ, M., REYNOLDS, J. J. and MELCHOLAM, R. (1964) *Tetrahedron Letters* **3065**; *Idem* (1965) *J. Org. Chem.* **30**, 4234.

²² DUGAN, J. J. and DE MAYO, P. (1965) *Can. J. Chem.* **43**, 2033; SHIMIZU, Y. and PELLTIER, S. W. (1966) *J. Am. Chem. Soc.* **88**, 1544.

²³ KUBOTA, T. and HINO, H. (1968) *Tetrahedron* **24**, 675.

internal microflora of rat, has been shown to hydrolyse numerous saponins from different plants thereby showing a low degree of specificity, but it is promising enough to be used as a tool for obtaining genuine sapogenins.²⁴



Acerotin	22-OAc, 21-OCCH=CHCH=CHBu, Δ^{12} , 28-COOH
Acerocin	22-OAc, 21-OCCH=CHCH=CHSec Bu, Δ^{12} , 28-COOH
Anagalligenone	23-OH, 16-oxo, 13 β $\rightarrow$ 28 epoxy
Barringtogenol B	21 β , 22 α , 28-OH, 16 α -anageloyloxy, Δ^{12}
Camelliagenin A	16 α , 22 α , 28-OH, Δ^{12}
Camelliagenin B	16 α , 22 α , 28-OH, 23-CHO, Δ^{12}
Camelliagenin C	16 α , 22 α , 23, 28-OH, Δ^{12}
Caccigenin	2 α , 21 β , 23-OH, Δ^{12} , 28-COOH
Caccigenin lactone	2 α , 23-OH, 21 $\rightarrow$ 28 lactone, Δ^{12}
16-Desoxy barringtogenol C	21 β , 22 α , 28-OH, Δ^{12}
Gymnemagenin	16 β , 21 β , 22 α , 23, 28-OH, Δ^{12}
Isothankunic acid	5, 6, 23-OH, Δ^{12} , 28-COOH
Platicodigenin	2 β , 16 α , 23, 24-OH; Δ^{12} , 28-COOH
Polygalacic acid	2 β , 16 α , 23-OH, Δ^{12} , 28-COOH
Proceric acid	21 β -OH, Δ^{12} , 28-COOH
Protoprimulagenin A	16 α -OH, 13 $\rightarrow$ 28 epoxy
Sapogenin K	2, 21-OH, Δ^{12} , 28-COOH
Spergulagenic acid	Δ^{12} , 28, 30-COOH
Serratagenic acid	Δ^{12} , 28, 29-COOH
Saikogenin A	16 β , 23, 28-OH, $\Delta^{11, 13(18)}$
Saikogenin B	16 β , 28-OH, $\Delta^{9(11), 12}$
Saikogenin C	16 β , 28-OH, $\Delta^{11, 13(18)}$
Saikogenin D	16 α , 23, 28-OH, $\Delta^{11, 13(18)}$
Saikogenin E	16 β , -OH, 13 $\rightarrow$ 28 epoxy, $\Delta^{11(12)}$
Saikogenin F	16 β , 23-OH, 13 β $\rightarrow$ 28 epoxy, $\Delta^{11(12)}$
Saikogenin G	16 α , 23-OH 13 β $\rightarrow$ 28 epoxy, $\Delta^{11(12)}$
Saniculagenin	3,3-dimethyl acrylic ester of R ₁ barrigenol
Sapindic acid	3 ξ $\rightarrow$ 23 epoxy, 28-COOH, Δ^{12}
Sapogenol T-A	15 α , 16 α , 22 α , 28-OH, 21 β -anageloxoy, Δ^{12}
Sapogenol T-B	16 α , 22 α , 28-OH, 21 β -anageloxoy, Δ^{12}
Theasapogenol A	16 α , 21 β , 22 α , 23, 28-OH, Δ^{12}
Theasapogenol E	16 α , 21 β , 22 α , 28-OH, 23-CHO, Δ^{12}

FIG 1 NEW SAPOGENINS OF THE β -AMYRIN GROUP

A novel cleavage method for splitting the glycosidic linkage of triterpene saponins by using soil bacteria giving the genuine sapogenins in good yield, has been reported in the cases of saponins from *Senega* and *Panax*,²⁵ *Aesculus* (horse chestnut),²⁶ *Styrax* and *Primula* spp. and *Lysimachia mauritiana*.²⁷

²⁴ GESTETNER, B., BIRK, Y. and TENCER, Y. (1968) *J Agr Food Chem* **16**, 1031

²⁵ YOSIOKA, I., FUJIO, M., OSAMURA, M. and KITAGAWA, I. (1966) *Tetrahedron Letters* 6303

²⁶ YOSIOKA, I., IMAI, K. and KITAGAWA, I. (1967) *Tetrahedron* **23**, 2577

²⁷ KITAGAWA, I., MATSUDA, A. and YOSIOKA, I. (1968) *Tetrahedron Letters* 5377, YOSIOKA, I., SAUOH, S. and KITAGAWA, I. (1972) *Chem Pharm Bull* **20**, 564

The structure of the sugar moiety is generally determined by mild acidic or alkaline hydrolysis of the permethylated glycosides. The individual methylated sugars are then identified either by direct chromatographic comparison or by gas chromatography (GLC). Mass spectrometry is also emerging as an invaluable tool in these investigations.

Some unusual and interesting features found in certain saponins are the presence of glucose on C₂ and C₆ of genins, in addition to C₃, in *Bassia* and *Dianthus* saponins respectively, identification of 3-*O*-methylglucose from stichoposides A and C, 3-*O*-acetylglucose in sanguisorbin E, α -D-glucose in calendulose and glucose in the furanose form in patrinosides C and D.

Measurements of UV, IR, NMR and MS are being used now as routine techniques in stereochemical and structural determination studies. These aspects have been covered in a recent review on the triterpenoids.^{2,7,8} By far the largest number of newly reported saponins belong to β -amyrin group, as was also found in the earlier survey.¹ Thus, out of the 54 new genins, 32 belong to this group, two to the hopane group and 19 to lanostane group amongst which are included 15 holothurinogenins of animal origin. The new saponins characterized during this period are given in Figs. 1-4.

It may be mentioned that the structure of platycodigenin has been determined by X-ray crystallography of its bromolactone and it is the first example of a naturally-occurring triterpene having geminal hydroxymethyl groups at C-4 in ring A.

The hopane group of genins were not encountered in nature so far and its first representatives, mollugogenol A (3 β , 6 α , 16 β , 22-tetrahydroxy isohopane), and Mollugogenol B (3 β , 6 α dihydroxy-hop-15,17 (21) diene) have been reported. An interesting C₃₀-Nortriterpenoid, eupteleogenin has also been found during this period (Fig. 2).

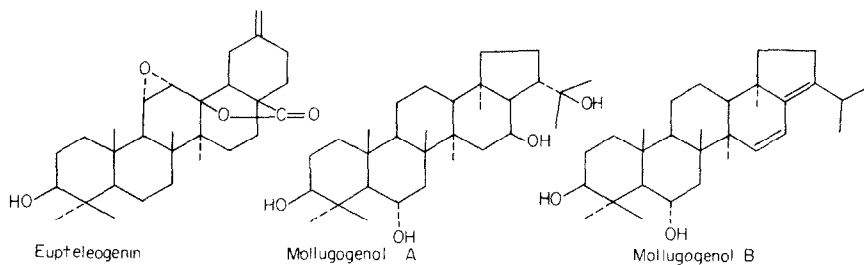


FIG. 2. HOPANE AND NORTRITERPENOID SAPONINS.

In the tetracyclic triterpenoid group, only four genins, aglycon-I, 16-acetyl tumulosic acid, its dehydroproduct and dahurinol have been obtained (Fig. 3).

Sea cucumbers secrete, presumably as a defence mechanism, water-soluble glycosides, holothurins, which possess greater hemolytic activity than saponins of plant origin. Holothurin itself is a mixture of glycoside sulphates and under varying conditions of hydrolysis, more than twelve genins have been isolated. The genins containing 7,8,9,11 heteroannular diene system and methoxyl group are artefacts because these groups are absent in parent holothurin and the structure of genuine aglycone moiety is still to be elaborated. These genins contain an unusual lactone ring (18 $\rightarrow$ 20) and belong to lanostane group. Stichopogenins A₂ and A₄ differ from holothurinogenin in the position of double bonds (Fig. 4).

^{2,7} KULSHRESHTHA, M. J., KULSHRESHTHA, D. K. and RASTOGI, R. P. (1972) *Phytochemistry* **11**, 2369.

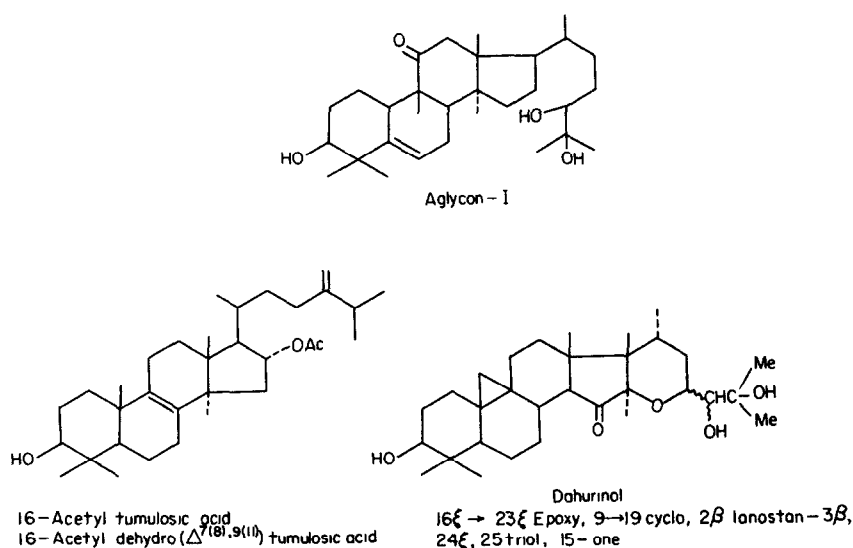


FIG 3 SAPOGENINS OF THE TETRACYCLIC TRITERPANOID GROUP

LIST OF SAPONINS CHARACTERIZED FROM 1966-72

The survey has been divided into five sections based on the isolation and characterization of saponins and their sapogenins, including the configuration of the carbohydrate moieties. The sugar components, wherever identified, are included (Table 1 A-E).

TABLE 1A SAPONINS WHOSE CHEMISTRY HAS BEEN COMPLETELY ELABORATED INCLUDING THE CONFIGURATION OF THE CARBOHYDRATE CHAIN

Plant	Saponin m p °C	Structure	Ref.
1 <i>Achyranthes aspera</i>	Saponin A	Oleanolic acid, α -L-Rha(1 $\rightarrow$ 4) β -D-glu (1 $\rightarrow$ 4) β -D-glu.A-C ₃	28
	Saponin B	β -D-Gal-C ₂₈ ester of saponin A	
2 <i>Akebia quinata</i> Decne	Saponin A 228-9 d	Same as scabioside A	29
	Saponin B 249-50 d	Hederagenin, β -D-Xyl(1 $\rightarrow$ 2)- α -L-ara-C ₃	
	Saponin C 248-50 d	Hederagenin, β -D-Glu(1 $\rightarrow$ 2)- α -L-ara-C ₃	
3 <i>Bassia latifolia</i> Roxb	Saponin A	Bassic acid, β -D-Glu-C ₂ ; β -D-Glu-C ₃	30
	Saponin B	Rha(1 $\rightarrow$ 4)xyl(1 $\rightarrow$ 2)-ara-C ₂₈ ester of saponin A	
4 <i>Beta vulgaris</i>	Glycoside A 248-50	Methyl oleanolate, β -D-Glu-C ₃	31
	Glycoside B 242-4	Oleanolic acid, β -D-Glu-C ₃	
	Glycoside C	Oleanolic acid, β -D-Glu A-C ₃	

TABLE 1A (cont.)

Plant	Saponin m.p. °C	Structure	Ref.
5 <i>Bupleurum falcatum</i> L.	Saikosaponin A 225–32	Saikogenin I β -D-Glu(1 $\rightarrow$ 3) β -D-fuc-C ₃	32
	Saikosaponin C 202–10	Saikogenin L β -D-Glu(1 $\rightarrow$ 6)-[α -L-rha(1 $\rightarrow$ 4)]- β -D-glu-C ₃	
	Saikosaponin D 212–18	Saikogenin G β -D-Glu(1 $\rightarrow$ 3)- β -D-fuc-C ₃	
6 <i>Calendula officinalis</i>	Calendulose A	Oleanolic acid β -D-Gal(1 $\rightarrow$ 4)- β -D-glu-C ₃	33
	Calendulose B	Oleanolic acid β -D-Gal(1 $\rightarrow$ 4)- β -D-glu-C ₃	
7 <i>Caltha silvestris</i>	Calcoside D 248–53	Hederagenin β -D-Glu(1 $\rightarrow$ 2) α -L-ara-C ₃	34
8 <i>Caulophyllum robustum</i>	Saponin I 224–6 d	Hederagenin α -L-ara-C ₃	35
	Saponin II 252–5 d	Same as Calcoside D	
9 <i>Chenopodium anthelminticum</i>	Saponin I	Echinocystic acid β -D-Xyl(1 $\rightarrow$ 6) β -D-glu(1 $\rightarrow$ 2) α -L-ara-C ₃	36
	Saponin II	β -D-Glu(1 $\rightarrow$ 4) α -L-rha-C ₂₈ ester of saponin I	
10 <i>Clematis mandshurica</i>	Clematoside A	Oleanolic acid D-Glu(1 $\rightarrow$ 4)-D-xyl(1 $\rightarrow$ 2) α -L-ara(1 $\rightarrow$ 2)-L-ara(1 $\rightarrow$ 4)-L-rha-C ₃ L-Rha(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 4)-D-glu-C ₂₈	37
	Clematoside A	Oleanolic acid D-Glu(1 $\rightarrow$ 4)-D-xyl(1 $\rightarrow$ 2)-L-ara(1 $\rightarrow$ 4)-L-rha-C ₃	
	Clematoside B	Oleanolic acid D-Glu(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 4)-D-xyl(1 $\rightarrow$ 2) α -L-ara(1 $\rightarrow$ 2)-L-rha(1 $\rightarrow$ 4)-L-rha-C ₃ L-Rha(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 6)-D-glu-C ₂₈	
	Clematoside C	Oleanolic acid L-Rha(1 $\rightarrow$ 6)-D-glu(1 $\rightarrow$ 6)-D-glu(1 $\rightarrow$ 4)-D-xyl(1 $\rightarrow$ 2)-L-ara(1 $\rightarrow$ 4)-L-rha-C ₃ L-Rha(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 4)-D-glu(1 $\rightarrow$ 6)-D-glu-C ₂₈	
11 <i>Dianthus deltoides</i>	Dianthoside A 220–5	Gypsogenic acid β -D-Glu-C ₃	38
	Dianthoside B 229–34	Gypsogenic acid β -D-Glu-C ₃	
12 <i>Dipsacus azureus</i>	Dipsacoside B	Hederagenin α -L-Rha(1 $\rightarrow$ 2) α -L-ara-C ₃ β -D-Glu(1 $\rightarrow$ 6)- β -D-glu-C ₂₈	39

TABLE 1A (cont)

Plant	Saponin m p °C	Structure	Ref
13 <i>Gypsophylla trichotoma</i>	Trichoside A	Gypsogenin, β -D-Glu(1 $\rightarrow$ 3)- β -D- glu A-C ₃ , β -D-Gal-C ₂₈	40
14 <i>Hedera caucasigena</i>	Hedera- caucaside B	Oleanolic acid, L-Rha(1 $\rightarrow$ 4) β -D-glu (1 $\rightarrow$ 6) β -D-glu-C ₃	41
15 <i>H. colchica</i>	Hedera- colchiside E	Oleanolic acid, L-Rha-L-ara(D-glu)-C ₃ , L-Rha(1 $\rightarrow$ 4) β -D-glu (1 $\rightarrow$ 6) β -D-glu-C ₂₈	42
	Saponin	Oleanolic acid D-Glu(1 $\rightarrow$ 4)[L-rha(1 $\rightarrow$ 5)]- *2 deoxyhex ² -C ₃ , *3-Deoxyhex ³ (1 $\rightarrow$ 4) β -D-glu(1 $\rightarrow$ 6)- β -D-glu-C ₂₈	
16 <i>H. pastuchovi</i>	Pastuchoside B	Hederagenin, α -D-Glu(1 $\rightarrow$ 4)- α - D-glu(1 $\rightarrow$ 3)- α -L-rha-C ₃	43
	Pastuchoside C	Hederagenin, α -L-Rha-C ₃	
17 <i>Helianthus annuus</i> (Sun flower)	Helianthoside B	Cochalic acid, D-Xyl-(1 $\rightarrow$ 4)-L-rha-C ₃ , D-Glu(1 $\rightarrow$ 4)-L-rha(1 $\rightarrow$ 2) L-ara-C ₂₈	44
	Helianthoside C	Cochalic acid, L-Rha(1 $\rightarrow$ 4)-L-rha(1 $\rightarrow$ 3) [D-xyl(1 $\rightarrow$ 6)]-D-glu-C ₃ , D-Glu(1 $\rightarrow$ 4)-L-rha(1 $\rightarrow$ 2)- L-ara-C ₂₈	
18 <i>Herniaria glabra</i>	Saponin I	Medicagenic acid, β -D-Glu(1 $\rightarrow$ 6)- β -D- glu-C ₂₈	45
	Saponin II	Medicagenic acid, β -D-Glu(1 $\rightarrow$ 4)[β -D-fuc (1 $\rightarrow$ 2)]- α -L-rha-C ₂₈	
19 <i>Koeleruteria paniculata</i> Laxm	Saponin A	Same as scabioside A	46
	Saponin B	Gypsogenin, α -L-Rha(1 $\rightarrow$ 3) α -L-ara (1 $\rightarrow$ 4)[β -D-gal(1 $\rightarrow$ 3)]-D- gal A-C ₃	
20 <i>Leontice eversmanni</i>	Leontoside A 276-8	Hederagenin, α -L-Ara-C ₃	47
	Leontoside B 242-4	Hederagenin, β -D-Glu(1 $\rightarrow$ 4) α -L- ara-C ₃	
	Leontoside C 222-4	Hederagenin, β -D-Glu(1 $\rightarrow$ 3) [β -D-glu(1 $\rightarrow$ 4)]- α -L- ara-C ₃	
	Leontoside D 202-4	Hederagenin, β -D-Glu(1 $\rightarrow$ 4) α -L-ara-C ₃ , α -L-Rha(1 $\rightarrow$ 4) β -D- glu(1 $\rightarrow$ 6)- β -D-glu-C ₂₈	

TABLE 1A (cont.)

Plant	Saponin mp °C	Structure	Ref
	Leontoside E 202-4	Hederagenin, at C-3 As in leonto- side C at C-28 As in leontoside D	
21 <i>Medicago sativa</i>	Saponin	Medicagenic acid β -D-Glu(1 $\rightarrow$ 6)- β -D- glu(1 $\rightarrow$ 3)- β -D-glu-C ₃	48
22 <i>Patrinia intermedia</i>	Patrinostide D	Oleanolic acid β -D-Glu(1 $\rightarrow$ 3)-[β -D-xyl (1 $\rightarrow$ 2)]- α -L-rha(1 $\rightarrow$ 4) β -D-xyl-C ₃ β -D-Glu(1 $\rightarrow$ 2) β -D-glucofuranosyl(1 $\rightarrow$ 4) β -D-glu-C ₂₈	49
	Patrinostide C	Oleanolic acid β -D-Xyl(1 $\rightarrow$ 2)- α -D-gluc- furanosyl(1 $\rightarrow$ 3)rha-C ₃ , at C-28 As in Patri- noside D	
23 <i>Patrinia scabiosifolia</i> Link	Scabioside A 216-9	Hederagenin, α -L-Ara-C ₃	50
	Scabioside B 210-2	Oleanolic acid, β -D-Glu(1 $\rightarrow$ 4) α -L-ara-C ₃	
	Scabioside C 216-8	Hederagenin, β -D-Glu(1 $\rightarrow$ 4) α -L-ara-C ₃	
	Scabioside D 224-6	Oleanolic acid, β -D-Glu(1 $\rightarrow$ 4) α -L-ara-C ₃ , β -D-Xyl-C ₂₈	
	Scabioside E	Oleanolic acid, β -D-Glu(1 $\rightarrow$ 4) α -L- ara-C ₃ , α -L-Rha(1 $\rightarrow$ 4)- β -D-xyl-C ₂₈	
	Scabioside G 235-6	Oleanolic acid, β -D-Glu(1 $\rightarrow$ 4) α -L-ara-C ₃ , β -D-Glu(1 $\rightarrow$ 4) α -L-ara (1 $\rightarrow$ 3) α -L-rha(1 $\rightarrow$ 4) β -D- xyl-C ₂₈	
24 <i>Patrinia sibirica</i>	Sibiroside A 245-50	Hederagenin β -D-Glu-C ₃ α -L-Ara-C ₂₈	51
	Sibiroside C 245-8	Hederagenin, β -D-Glu(1 $\rightarrow$ 4) α -L- ara(1 $\rightarrow$ 4)- β -D-glu-C ₃ , D-Glu(1 $\rightarrow$ 4)- β -D-glu (1 $\rightarrow$ 4) α -L-ara-C ₂₈	
25 <i>Phaseolus vulgaris</i> L (Broad bean)	Phaseoloside D	Genin ⁹⁸ , β -D-3-Deoxyhex ⁹⁸ (1 $\rightarrow$ 2) β -D-glu(1 $\rightarrow$ 4)[α -L-rha- (1 $\rightarrow$ 6)] β -D-gal(1 $\rightarrow$ 2) α - L-ara(1 $\rightarrow$ 3) β -D-glu A-C ₃	52
	Phaseoloside E	Genin ⁹⁸ , β -D-glu(1 $\rightarrow$ 4) β -D-glu (1 $\rightarrow$ 4)- β -D-3-deoxyhex ⁹⁸ (1 $\rightarrow$ 2) β -D-glu(1 $\rightarrow$ 4) [α -L-rha(1 $\rightarrow$ 6)] β -D- gal(1 $\rightarrow$ 2) α -L-ara(1 $\rightarrow$ 3) β -D-glu A C ₃	

TABLE 1A (cont)

Plant	Saponin m p °C	Structure	Ref
26. <i>Putranjiva roxburghii</i>	Saponin A	Oleanolic acid, α -L-Rha(1 $\rightarrow$ 2) β -D- glu(1 $\rightarrow$ 3) β -D-glu A-C ₃	53
	Saponin B	Oleanolic acid, α -L-Rha(1 $\rightarrow$ 2)- β -D- glu(1 $\rightarrow$ 3)[β -D-xyl(1 $\rightarrow$ 4)] β -D-glu A-C ₃	
	Saponin C	β -D-Glu-C ₂₈ ester of saponin A	
	Saponin D	β -D-Glu-C ₂₈ ester of saponin B	
27. <i>Sanguisorba officinalis</i>	Sanguisorbin B 266-7	Ursolic acid, β -L-Ara-C ₃	54
	Sanguisorbin E 190-2	Ursolic acid, β -L-Ara-C ₃ , β -D-3- Acetyl-glu-C ₂₈	
28. <i>Sapindus mukorassi</i> (Chinese soap berry)	Sapindoside A 214-16	Hederagenin, α -L-Rha(1 $\rightarrow$ 2) α -L-ara-C ₃	55
	Sapindoside B 276-8	Hederagenin, β -D-Xyl(1 $\rightarrow$ 3) α -L- rha(1 $\rightarrow$ 2) α -L-ara-C ₃	
	Sapindoside C 235	Hederagenin, β -D-Glu(1 $\rightarrow$ 4) β -D-xyl (1 $\rightarrow$ 3) α -L-rha(1 $\rightarrow$ 2) α -L-ara-C ₃	
	Sapindoside E	Hederagenin, D-Xyl(1 $\rightarrow$ 3)-L-rha(1 $\rightarrow$ 2) L-ara-C ₃ , β -D-Glu(1 $\rightarrow$ 2) β -D-glu(1 $\rightarrow$ 4)[α -L-rha (1 $\rightarrow$ 6)] β -D-xyl(1 $\rightarrow$ 3) α -L-rha(1 $\rightarrow$ 2) α -L-ara-C ₂₈	
29. <i>Saponaria officinalis</i>	Saponaside A 132-4	Gypsogenin, β -D-Glu A-C ₃ , β -D-Glu(1 $\rightarrow$ 6) β -D- glu(1 $\rightarrow$ 3) β -D-glu-C ₂₈	56
	Saponaside D 241-4	Gypsogenin, α -L-Ara(1 $\rightarrow$ 4)[β -D-gal (1 $\rightarrow$ 2)] β -D-xyl(1 $\rightarrow$ 3) [α -L-rha(1 $\rightarrow$ 4)] β -D- glu A-C ₃ , β -D-Gal (1 $\rightarrow$ 4)[β -D-glu(1 $\rightarrow$ 2)] β -D-xyl(1 $\rightarrow$ 3)[α -L- rha(1 $\rightarrow$ 4)] β -D-fuc-C ₂₈	
30. <i>Xanthoceras spp</i>	Saponin	Gypsogenin, β -D-Ara(1 $\rightarrow$ 4) β -D-gal (1 $\rightarrow$ 6)[β -D-glu A(1 $\rightarrow$ 3)] β -D-glu-C ₃	57

Note All the reported sugars are present in pyranose form, unless otherwise mentioned
Abbreviations used Glu = glucose, Fru = fructose, Gal = galactose, Ara = arabinose, Fuc = fucose, Xyl = xylose, Rha = rhamnose, Glu A = glucuronic acid, gal A = galacturonic acid.

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TABLE 1B SAPONINS STATED TO BE PURE, WHOSE GENINS HAVE BEEN CHARACTERIZED

Plant	Saponin m p °C	Genins and Sugars	Ref
1 <i>Acacia concinna</i>	Acacinine 260-64	Acacic acid + glu + ara + rha	58
2 <i>Acer negundo</i>	Saponin P	Acerotin + acerocin + glu + ara	59
3 <i>Albizzia lebbek</i>	Lebbekanin A	Echinocystic acid + glu + ara + xyl + rha	60
	Lebbekanin B	Oleanolic acid + glu + ara + xyl + rha	
4 <i>A procera</i>	Proceranin A	Proceric acid + glu + gal + ara + xyl + fuc + rha	61
5 <i>Bassia longifolia</i>	Brassianin	Brassic acid + glu + ara + xyl + rha	62
6 <i>Calendula officinalis</i>	Calendulose A 260-62	Oleanolic acid + glu + gal	63
7 <i>Caulophyllum robustum</i>	Caulosides A, B	Hederagenin + glu + rha + ara	64
	Cauloside C	Hederagenin + glu + ara	
	Cauloside D	Hederagenin (?) + ara	
	Cauloside E	Hederagenin + ara	
8 <i>Centella asiatica</i>	Isothankun- unside	Isothankunic acid + glu + rha	65
9 <i>Chenopodium ambrosioides</i>	Chenopodo- side B 213-16	Echinocystic acid + rha + xyl + ara + glu A	66
10 <i>Dianthus deloides</i>	Dianthoside C 270-5	Gypsogenic acid + ara + fuc + glu A + gal + xyl + rha	38
11 <i>D superbus</i> var <i>longicalycinus</i>	Saponin D	Gypsogenin + glu	67
12 <i>Dipsacus azureus</i>	Dipsacoside A	Hederagenin + ara + rha	68
	Dipsacoside B	Hederagenin + glu + ara + rha	
13 <i>Eleutherococcus senticosus</i>	Senticosides A, B	Oleanolic acid + ara + rha	69
	Senticosides C, D, E, F	Oleanolic acid	
14 <i>Empetrum sibiricum</i>	Empetroside A 209-11	Ursolic acid + glu	12
	Empetroside B 293-5	Ursolic acid + glu	
	Empetroside C 185-8	Ursolic acid + glu + ara	
15 <i>Entada phaseoloides</i>	...	Entagenic acid + ara + xyl	70
16 <i>Euptelea polyandra</i>	Eupteleoside A	Eupteleogenin + glu + rha + ara	71
	Eupteleoside B	Monoacetate of eupteleoside A	
17 <i>Gleditschia triacanthos</i>	Triacanthoside C	Oleanolic acid + glu + ara + rha + xyl	72
18 <i>Glycyrrhiza echinata</i>	Saponin A	Macedonic acid + rha + glu A	73
	Saponin B	Echinatic acid + rha + glu A	
19 <i>Gypsophilla acutifolia</i>	Acutifolioside	Gypsogenin + glu A + gal + ara + xyl + rha	74
20 <i>G trichotoma</i>	Trichoside B	Gypsogenin + glu + gal + glu A	75
	Trichoside D	Gypsogenin + gal + ara + xyl + glu A + glu + rha + fuc	

TABLE 1B (CONT.)

Plant	Saponin m.p. °C	Genins and Sugars	Ref
21 <i>Hedera caucasicarpa</i> (Caucasian ivy)	Hederacataca- side B Hederacauca- sides A, C, D	Hederagenin + oleanolic acid + glu + rha Hederagenin + oleanolic acid	76
22 <i>H. pastuchovii</i>	Pastuchoside A (= Calopanax- saponin B) Pastuchosides B, D, E, F, G, H, I	Hederagenin + 2 glu + 2 rha + ara Oleanolic acid + hederagenin + glu + ara + rha	77
23 <i>Ladygina huchanica</i>	Ladyginoside C Ladyginoside D	Oleanolic acid + glu + ara + glu A Hederagenin + glu + glu A Primulagenin A + glu +	78
24 <i>Mysine africana</i>		2 rha + gal + glu A	79
25 <i>Olax subscoor- pionides</i>	-	Oleanolic acid + glu + xyl + rha	80
26 <i>Panax japonicum</i>	Saponin I 196.7 Saponin II 235 d Saponin III 230 d	Panaxadiol + glu + xyl Oleanolic acid + glu + ara + glu A Oleanolic acid + glu + glu A	81
27 <i>Phaseolus vulgaris</i>	Phaseoloside E 130	Soyasapogenol + glu + gal + ara + rha + glu A	82
28 <i>Platycodon grandiflorum</i>	Platycodoside C	Platycodogenin (C ₁₀ H ₄₆ O ₆) + polygallic acid	83
29 <i>Polygala senega</i>	Senegin II 247.8	Presenegenin + glu + gal + rha + xyl + fuc + 3,4 dimethoxy cinnamic acid	84
30 <i>Pulsatilla nigricans</i>	Pulsatoside	Hederagenin + ara + gal + glu + rha	85
31 <i>Putranjiva roxburghii</i>	Putranjivoside 215.7	Oleanolic acid + ara + rha + glu	86
32 <i>Sapindus emarginatus</i>	Emarginatoside 220.2	Hederagenin + glu + xyl + rha	87
33 <i>S. mukorossi</i>		Hederagenin + ara + xyl + glu A	88
	Mukorosside 155.6	Hederagenin + glu + xyl + ara + rha	10
	Saponoside D	Hederagenin + glu + xyl + rha + ara	89
34 <i>Saponaria officinalis</i>	Saponaroside 202.2 Saponarosides B, C	Gypsogenic acid + xyl Gypsogenin	90
35 <i>Spergularia arbuscula</i>	Espergularin 235.8 d	Oleanolic and echinocystic acids + gal + rha + xyl	91
36 <i>Ternstroemia japonica</i>	Saponin D Saponin C ₁	A ₁ - barrigenol + 2 rha + glu + gal + glu A Octadienoic acid ester of saponin D	92

Note: Saponin⁹³ from *Primula marcoviana* Bunge (calyx primrose) is identical to primula-saponin from *P. turkestanica*.

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TABLE 1C. SAPONINS ISOLATED AS PURE OR MIXTURES FROM GENUS-BASE SPECIES-BASE REFINED

Plant	Crude saponin m.p. °C	Genins (m.p. °C) and sugars	Ref.
1 <i>Acanthophyllum adenophorum</i>	212-18	Gypsogenin + gal + xyl + rha + ara + fuc	94
2 <i>Adina pilulifera</i>	Mixed saponins	Morolic, betulinic, quinovic and chincholic acids + an unidentified triterpene acid + glu + glu A	95
3 <i>Aegiceras corniculatum</i>	Mixed glycosides	Primulagenin + aegiceradienol + aegiceradiol	96
4 <i>Aesculus turbinata</i> Blume		Protoaescigenin + barringtongenol C (=aescinidin) + 16-desoxybarringtongenol C	97
5 <i>Alysicarpus amheimii</i> Tica	Massein	Feruloyl galactaric acid + ara + 3 glu	98
6 <i>Aprocera</i> Benth		Procera-machaerinic acids + ethyl machaerinic + macherinic acid (28 → 21) lactone + Procera-genin A m.p. 294 °C $C_{30}H_{46}O_4$ + Procera-genin B m.p. 255-8 °C $C_{30}H_{46}O_4$	99
7 <i>Anagallis arvensis</i>	Hetroside I	Anagalligenone	100
8 <i>Barringtonia acutangula</i> Gaertn	-	Barringtonenols B, C and D	101
9 <i>Blepharis edulis</i>	-	Lupeol	102
10 <i>Bryonia dioica</i>	Two glycosides	Aglycon-I + $C_{30}H_{48}O_3$ + glu + rha	103
11 <i>Bupleium falcatum</i> L.		Saikogenins A, B, C, D	104
<i>Bupleium</i> species	Saikoside Ib 213-17	Saikogenin A	
12 <i>Caccinia glauca</i>	230-57	Caccigenin + glu + rha	105
13 <i>Camellia japonica</i> L.	-	Camelliagenins A, B and C	106
14 <i>Cassipoupa</i> Thunb	-	Camellia sapogenols I and II + theasapogenols A, B & E (thea sapogenol B = barringtongenol C)	107
15 <i>Cimicifuga dahurica</i>		Dahurinol	108
16 <i>Clerodendron serratum</i>		Oleanolic, queretaronic and serratagenic acids	109
17 <i>Elyngium bieberschteinianum</i>		Barringerols R ₁ , A ₁	110
18 <i>Euphorbia cretica</i>	Fagonin	Oleanolic acid	111
19 <i>Gleditsia pteris</i>		Oleanolic, coehalic acids + glu + rha + xyl + ara	112
20 <i>Gymnema sylvestre</i> R. Br	-	Gymnemagenin	113
21 <i>Hedera ciliata</i>		Oleanolic acid + hederagenin	114
22 <i>H. helix</i>	amorph 195-200 Saponin lactone 225	Hederagenin	115

TABLE 1C (cont.)

Plant	Crude saponin m p °C	Genins (m p °C) and sugars	Ref
23 <i>Helianthus annuus</i>	Fraction A	Oleanolic, albigenic and echinocystic acids + glu + ara + rha + xyl (COOH of genin not free)	116
	Fraction B	Same as above (COOH of genin free)	
24 <i>Lysimachia clethroides</i>	—	Primulagenin A + dihydropriverogenin A	117
<i>L. japonica</i>	—		
25 <i>L. mauritiana</i>	—	Dihydropriverogenin A (=camelliagenin A) + Priverogenin B	118
26 <i>Medicago sativa</i> (Lucerne)	Lucerne saponin	Medicagenic acid + soyasapogenol A + aglycone + glu + xyl + ara	119
27 <i>Melianthus comosus</i>	—	Ellagic acid	120
28 <i>Mimulus elengi</i>	—	β -Amyrin + bassic acid + neutral genin	121
29 <i>Mollugo hirta</i>	—	Mollugogenols A and B	
30 <i>M. spargula</i>	—	Spergulagenic acid + neutral genin, C ₃₀ H ₅₀ O ₄	123
31 <i>Patrinia scabiosaeifolia</i>	Patrinosides mix	Oleanolic acid + xyl + ara	124
32 <i>Platycodon grandiflorum</i>	—	Polygalacic acid + platycodigenin	125
A De candolle			
33 <i>Pithecolobium dulce</i>	—	Oleanolic and echinocystic acids + xyl + ara + glu	126
34 <i>Pittosporum tobira</i> Ait	—	Barrigenol R ₁ + 21-O-anageloyl barrigenol-R ₁ (=sapogenol T-A) + 21-O-anageloyl barringtonol C (=sapogenol T-B)	127
35 <i>Polygala senega</i>	Saponins, A, B, C, D	Presenegenin + glu + rha + xyl + gal + fuc	128
36 <i>Primula sieboldi</i>	—	Protoprimulagenin A	129
37 <i>P. japonica</i>	—	Camelliagenin A (=dihydropriverogenin A)	129
38 <i>Sanicula europaea</i>	Saponins, A, B, C, D	Saniculagenin + genin B, m p 160-70 + genin C, m p 276-9 + glu + ara	130
39 <i>Sapindus laurifolius</i>	—	Methyl hederagenate + sapindic acid + methyl oleanolate + two triterpenes	131
40 <i>Stryphnodendron coriaceum</i>	—	Maslinic and machaerinic acids + sapogenin K	132
41 <i>Thea sinensis</i> L.	—	Barringtonol C (=theasapogenol B), theasapogenol A, camelliagenin C (=theasapogenol C), dihydropriverogenin A (=camelliagenin A = theasapogenol D), theasapogenol E	133
42 <i>Trametes dickinsii</i>	Mix of two trametosides	16-Acetyl tumulosic acid + 16-acetyl dehydrotumulosic acid + glu	134

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TABLE 1D SAPONINS, PURE OR CRUDE, WHOSE GENINS HAVE NOT BEEN CHARACTERIZED

Plant	Work done	Ref
1 <i>Acacia concinna</i>	Pure saponin, m p 252° → genin, C ₃₀ H ₄₈ O ₅ + glu	135
2 <i>Cucumaria fraudatrix</i>	Cucumarioside C → genin A ₃ (C ₃₀ H ₄₄ O ₅) + glu + xyl + rha	136
3 <i>Cyclamen vernal</i> <i>C ibericum</i> <i>C adzharicum</i> <i>C abchasicum</i>	Saponin, m p 252 → Aglycone + glu + ara	137
4 <i>C pseudoibericum</i>	Saponin, m p 270 → C ₃₀ H ₄₅ O ₆ + glu + xyl + ara	138
5 <i>Fagonia cretica</i>	Saponin → Fagogenin, m p 307° + C ₃₀ H ₄₆ O ₅ , m p 326° + glu + rha + ara	139
6 <i>Gleditschia spp</i> (Honey locust)	Pure saponin, m p 190° → aglycone + glu + fru	140
7 <i>G australis</i>	Saponin, C ₃₀ H ₄₈ O ₄ , m p 296°	141
8 <i>G triacanthos</i>	Triacanthosides, A, B and D	142
9 <i>Gypsophilla trichotoma</i>	Trichosides A and C	143
10 <i>Hedera pastuchovi</i>	Pastuchoside C, m p 254° → C ₃₀ H ₄₈ O ₄ + hederagenin + rha	144
11 <i>Hydrocotyle vulgaris</i>	Saponin → Cotylegenins A, B, C, D & E + glu + ara	145
12 <i>Ladygina bucharica</i>	Ladyginosides A, B & E	78
13 <i>Lyonia ovalifolia</i> var <i>elliptica</i>	Ovalifolioside, C ₃₅ H ₅₈ O ₇ → ovalifoligenin, C ₃₀ H ₅₀ O ₃ + ara	146
14 <i>Panax ginseng</i>	Panaxoside B' → 3 glu Panaxoside C → 2 glu + rha	147
15 <i>Polemonium coeruleum</i>	Polemonioside B, m p 232° → Gal Polemonioside C, m p 241° → Ara + gal	148
16 <i>Polemonium acutifolium</i> <i>P pauciflorum</i> <i>P occidentale</i>	Three saponins → 2 aglycones + gal + ara	149
17 <i>Sanguisorba officinalis</i>	Sanguisorbin A, m p 272° → genin, m p 242° + ara Waremokonin, m p. 204° → genin + ara + glu + acetic acid	150
18 <i>Securigera securidaca</i>	Crude saponin → glu + gal + rha	151
19 <i>Sophora prodani</i>	Saponin, m p 220°	152
20 <i>Valeriana officinalis</i>	Valeroside A, m p 298° → genin + glu	153
21 <i>Zygophyllum fabago</i>	Zygophylosides A, B, C, D & E → genin, m p 305° + gal + glu + ara	154

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TABLE 1E. SAPONINS OF ANIMAL ORIGIN FROM HOLOTHURIANS (SEA-CUCUMBERS)—PHYLUM *Echinodermata*

Name	Saponin m.p. °C	Genins and sugars	Ref.
1 <i>Cucumaria frandatrix</i>	Cucumarioside C	Genin A ₃ (C ₃₀ H ₄₄ O ₈) + glu + xyl + rha	155
2 <i>Actinopyga aqassizi</i>	Holothurin A	22,25-oxidoholothurinogenin*, 17-deoxy-22,25-oxidoholothurinogenin + xyl + quinovose	155a
3 <i>Holothuria vagabunda</i> <i>H. lubrica</i> <i>H. leucopilota</i>	Holothurin B 213–23 d	12β-methoxy-7,8 dihydroholothurinogenin (neo-holothurinogenins)	
<i>H. poli</i>		Griseogenin, praslinogenin, teinaygenin, seychellogenin, koellikerigenin	155b
4 <i>Bohadschia koellikeri</i>			
5 <i>Stichopus japonicus</i> Selenka	Stichoposide A 215–17	Stichopogenin A ₂ and A ₁ + glu + xyl + 3-O-methyl glu	156
	Stichoposide C	Stichopogenins + glu + gal + xyl + 3-O-methyl glu	

* The various holothurinogenins are artefacts produced during hydrolysis of the saponins

It may be of interest to mention that asteroids (sea stars), another class of phylum *Echinodermata*, also elaborate saponins which belong to steroid group and are highly toxic.^{156,1}

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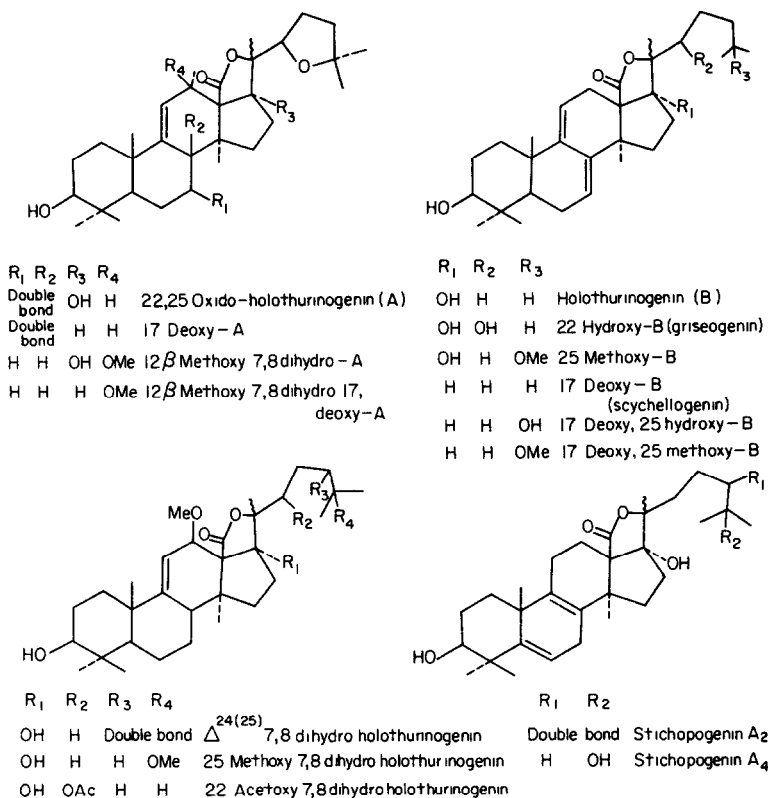


FIG 4 SAPOGENINS OF THE SEACUCUMBERS

BIOLOGICAL ACTION AND USES

The significance of saponins in pharmacy with special reference to surface activity, complex formation, hemolysis, toxicity and various therapeutic effects has been reviewed.¹⁵⁷ The relationship between the presence of saponins in leguminous plants and the appearance of tympanites in ruminants and the present knowledge of the depressive action of leguminous saponins on the growth of chicks and egg laying have also been reviewed.¹⁵⁸

The hemolytic activity of the saponins have been found to be very variable. The water soluble saponin from *Hedera helix* leaves is devoid of hemolytic properties whereas water-insoluble saponin lactone from the same source and the magnesium salt of its corresponding hydroxy acid showed a potent activity.¹¹⁵ The hemolytic index of pure saponin from *Calyx primrose* was found to be 20.¹⁵⁹

¹⁵⁷ GERLACH, I H (1967) *Pharm Zentralh Deut* **106**, 573; (1967) *Chem Abstr* **67**, 111362 x; ZVAVING, J H. (1971) *Pharm Weekbl* **106**, 629

¹⁵⁸ MAYMORE, B (1962) *Ann Ist Sper Zootec Roma* **9**, 429, (1966) *Chem Abstr* **65**, 20575 f

¹⁵⁹ ASOEVA, E Z, DAUKSHA, A D, DENISOVA, E K and MURAV'eva, D A (1967) *Vch Zap Pyatigorsk Farm Inst* **6**, 8, From Ref *Zh Biol Khim* (1968) Abstract No 18F 1049, (1969) *Chem Abstr* **70**, 103719 r

Lucerne saponin showed strong hemolytic activity and depressed growth of the beetle (*Tribolium castaneum*) larvae but its inhibitory effect could be reversed when cholesterol was incorporated in diet. Germination of cotton seeds was inhibited in soils on which lucerne has been grown or on soils supplemented with lucerne meal from tops and roots¹⁶⁰

The structural specificity of saponin hemolysis has been studied by testing a large number of saponins and triterpene aglycones using cattle erythrocytes *in vitro*. It has been shown that the requirements for optimum activity are a polar grouping in ring A and a moderately polar grouping in ring D or E. The compounds containing 16 α -OH or 16-keto group together with 3 β -OH had the highest hemolytic index. The distance of 10.5 Å between 3 β -OH and 16 α -OH is of special significance. Acylation of the either hydroxyl group resulted in loss of activity. The sugar chains (in glycosides) had a certain influence on the hemolytic activity. A polar group in ring D and/or E, such as sugar chain or a number of OH groups, induced inactivation¹⁶¹

A significant antimitotic activity has been observed in the glycosides and their magnesium salts from *Hedera helix*. These substances inhibited the growth of cells without destroying them and can be used for inhibiting the growth of benign or malignant tumors and for combating protozoa, in particular *Trichomonas*. Although they widely differ in hemolytic activity, they are relatively non-toxic for mice, rats and guinea pigs and a process patent has been taken for their production.¹¹⁵ The saponin fraction of *Acer negundo* and *Cyclamen persicum* showed inhibitory activity against Walker 256 tumor in rats.¹⁶² The saponins isolated from *Myrsine africana* and *Entada phaseoloides*, have shown significant inhibitory activity against the Walker 256 in rats.⁷⁰⁻⁷⁹

Some saponins have shown prominent anti-inflammatory activity. The mixture of saikosides from *Bupleurum falcatum* is effective (400 mg/kg) for treatment of rat edema.¹⁰⁴ The crude saponin of *Thea sinensis* showed pronounced antiexudative and anti-inflammatory properties. It normalized the capillary permeability which is disturbed in the initial stage of inflammation.¹⁶³ The saponins from *Mimusops manikara* and *Pithecolobium dulce*, hederagenin and sodium nimbinat showed anti-inflammatory activity against carragenin-induced edema and formaldehyde-induced arthritis in rats; ED₅₀, LD₅₀ and structure activity relations have been described.¹⁶⁴ A gypsogenin saponin from *Momordica cochinchinensis* had LD₅₀ 37 mg/kg in mice, inhibited rat paw edema and intestinal mobility of rabbit duodenum. The saponin at 3×10^{-4} g/ml potentiated the contractile response of guinea pig ileum to acetyl choline (10^{-9} g/ml) and abolished the anti-acetylcholine action of papavarine (2×10^{-5} g/ml).¹⁶⁵

The antifungal and antimicrobial effects of escin, *Gypsophylla* saponin, *Quillaja* saponin, primulic acid and ammonium glycyrrhizate upon the hyphal growth of *Claviceps purpurea*.

¹⁶⁰ SHANY, S., BIRK, Y., GESTITNER, B. and BONDI, A. (1970) *J. Sci. Food Agr.* **21**, 131. GESTITNER, B., SHANY, S., TINCER, Y., BIRK, Y. and BONDI, A. (1970) *ibid.* **21**, 502.

¹⁶¹ SCHLOESSER, E. and WULF, G. (1969) *Z. Naturforsch. B* **24**, 1284.

¹⁶² KUPCHAN, S. M., HEMINGWAY, R. J., KNOX, J. R., BARBOUTIS, S. J., WERNER, D. and BARBOUTIS, M. A. (1967) *J. Pharm. Sci.* **56**, 603.

¹⁶³ VOGL, G., MARIK, M. L. and OERTNER, R. (1968) *Arzneim-Forsch.* **18**, 1466, (1969) *Chem. Abstr.* **70**, 36279 g.

¹⁶⁴ BHARGAVA, K. P., GUPTA, M. B., GUPTA, G. P. and MISHRA, C. R. (1970) *Indian J. Med. Res.* **58**, 724.

¹⁶⁵ KITAHARA, K., SATO, M., MURAKAMI, T. and YAMAGISHI, T. (1971) *Yakugaku Zasshi* **91**, 174. (1971) *Chem. Abstr.* **74**, 139209.

Piricularia oryzae, *Trichothecium roseum* and *Polyporus versicolor* were determined *in vitro*. Only escin and primulic acid at 50–400 µg/ml inhibited hyphal growth.¹⁶⁶

The fungistatic action of 19 saponins and saponin fractions were tested on 15 species of fungi, mostly plant pathogens. *Sclerotinia fructicola*, *C. purpurea*, *T. roseum*, *P. oryzae* and *Fomes officinalis* were most sensitive to the saponins, whereas *Botrytis allii*, *Alternaria solani*, *Coniophora cerebella*, *Rhizoctonia solani*, *Schizophyllum commune* and 3 *Fusarium* species were quite resistant. *T. roseum* was sensitive to saponins but was resistant to steroid saponins with double bond at C-5 as in α -solanine etc. *Polystictus* was completely inhibited by steroid saponins but only weakly effected by triterpene saponins whereas the reverse was true for *Coniophora* and *Aspergillus*.¹⁶⁷ The glycoside of *Anagallis arvensis* revealed antifungal action and inhibited germination of linseed. Its LD₅₀ in mice was 675 mg/kg *per os* and 30.4 mg/kg by *s.c.*¹⁶⁸ Eupteleoside A and B from *Euptelea polyandra* also possess intense anti-microbial activity¹⁶⁹ against *Saccharomyces cerevisiae* and *Piricularia oryzae* as well as some phytopathogenic fungi. A synergistic effect of A and B is also described. In another study avenacin, the resistance factor in oat roots, and escin were found to inhibit the growth of various fungi including *Ophiobolus graminis* and *Neurospora crassa*. These saponins were found to cause a rapid loss of inorganic ions—K from the mycelium of the former and Mg from latter—in the phosphate buffer. Similar action was noted from the mycelia suspended in acetate buffer. The saponins also inhibited the uptake of K and Mg in fungal mycelium both in phosphate and succinate buffers.¹⁷⁰

The effect of *Sapindus mukorossi* saponins (a mixture of five glycosides of hederagenin) on blood pressure and cholesterol content in mice has been studied. The saponins showed LD₅₀ (mg/kg) *per os* 1625, *s.c.* 650, *iv* and *ip* 276 in mice. In normal rabbits saponins administered *per os* (0.04 mg/kg) did not change arterial pressure. If injected (*s.c.*) at the same dose they decreased blood pressure by 25% during first 2–3 hr but blood cholesterol content remained unchanged. In rabbits with experimental atherosclerosis induced by feeding cholesterol rich diet, saponins administered *per os* for 30–50 days, beginning from the 40th day decreased the cholesterol level from 292–248 to 53–110 mg%.¹⁷¹

The action of *Cyclamen ibericum* saponin in tests on cats, frogs, isolated frog hearts, rabbits and monkeys, showed positive inotropic and negative chromotropic action but no significant effect on arterial pressure and respiration. It also exhibited vasoconstrictive effect in isolated rabbit ears and increased permeability of dermal capillaries in rats.¹⁷² The effect of *Leontice eversmanni* saponin and glycyrrhizic acid on experimental atherosclerosis was evaluated on three groups of rabbits which were given 0.3 g/kg of cholesterol daily for 60 days. In addition, group 2 received 10 mg/kg saponin and group 3 received 15 mg/kg of K-glycyrrhizate. After 60 days animals were killed and the degree of atherosclerosis was evaluated. The protective action against atherosclerotic changes was more pronounced with saponin than with glycyrrhizic acid. The cholesterol blood levels in the respective

¹⁶⁶ WOLTERS, B (1966) *Naturwiss* **53**, 253

¹⁶⁷ WOLTERS, B (1968) *Planta* **79**, 77

¹⁶⁸ STARON, T, ALLARD, C, HEITZ, S and BILLET, D (1969) *Phytat-Phytopharm* **18**, 161, (1971) *Chem Abstr* **75**, 59769 j

¹⁶⁹ GOTO, M, MURATA, T, IMAI, S and FUJIOKA, S (1970) *Yakugaku Zasshi* **90**, 736, (1970) *Chem Abstr* **73**, 84655 k

¹⁷⁰ OLSEN, R A (1971) *Physiol Plant*, **24**, 534, (1971) *ibid* **25**, 204, 503

¹⁷¹ EFIMOVA, T G, PIVNENKO, G P, KUTSEVICH, V A and ZIKOVA, N YA (1966) *Farm Zh (Kiet)* **21**, 45, (1967) *Chem Abstr* **66**, 93780 e

¹⁷² GLADKIKH, A S (1966) *Farmakol Toksikol* **29**, 559, (1966) *Chem Abstr* **66**, 9774 s

groups were as follows: Group (1) 600–775, (2) 400–500; (3) 500–600 mg¹⁷³. In addition the cholesterol: lecithin ratio was reduced by saponin and glycyrrhizic acid¹⁷³

Aralosides A, B and C of *Aralia manschurica* upon intravenous injection at 10 mg/kg markedly stimulated the electrical activity of the rabbit brain and increased the processes of nerve cell excitability and lability¹⁷⁴

The acid and neutral saponins of *Patrinia intermedia* when administered *s.c.* to mice at 10–100 mg/kg, reduced the spontaneous motor (CNS depressant) activity, prolonged hexanal (70 mg/kg, *i.p.*) induced sleep and in rats, at the same dose, inhibited conditioned reflex reaction. Prior administration of the glycoside (0.2–0.3 g/kg) reduced the convulsive action of strychnine (1 mg/kg *s.c.*) and increased the life span.¹⁷⁵ The saponins of *Centella asiatica* exhibited sedative action in rat and mice and the mode of action appeared to be mainly on the cholinergic mechanism in CNS.¹⁷⁶

The comparative study of pharmacological properties of *Bupleurum longeradiatum* var *breviradiatum* and *B. falcatum* saponins has been made. *Per os* administration of 1.0–3.0 g/kg of the saponins was not toxic and the analgesic effect of the former was twice that of the latter. Both the saponins (500 mg/kg *per os*) prolonged sleeping time induced by hexobarbital sodium in mice. They also decreased the blood pressure of rabbit (10–25 mg/kg), caused cardiac inhibition of isolated frog heart and guinea pig arterial preparations and increased the peristalsis of isolated rabbit intestine ($1-2 \times 10^{-4}$ g/ml). Rat paw edema induced by dextran, 5-HT and croton oil was suppressed by *per os* administration of 600 mg/kg of both the saponins whereas inhibition of pleural exudation after 0.1% Evans blue and 0.2% gum arabic, was only slight. Thus, there was no difference in the actions of the two saponins except in the analgesic effect¹⁷⁷

The termite and fungal resistance of the ancient woods in temples have been found to be due to the presence of saponins.¹⁷⁸ Saponins, aglycones have been found to strongly inhibit the voracity of the termites.¹⁷⁹ The antitermitic behaviour of the *Temstroemia japonica* wood was shown to be due to saponins C₁, C₂ and D which were active in the order C > D > C₂⁹²

A process has been patented for an antigenic vaccine with a saponin adjuvant for immunizing vertebrates against protozoal diseases. The antigenicity of an antigen preparation obtained from the trypanosomes of protozoa of *Trypanosoma* species, has been enhanced by admixture with the aqueous solution of a saponin preparation, obtained from the bark of *Quillaja saponaria* or the root of *Saponaria officinalis*. The resulting antigen preparation was to immunize against diseases caused by pathogenic protozoa as Chagas' disease and trypanosomiasis¹⁸⁰

An improved process for the commercial production of three active glycosides, m.p. 260–270, 215–220 and a syrup respectively, from *Panax ginseng*, has been developed and

¹⁷³ EVODOKIMOVA, N. I. and KANILOV, I. K. (1967) *Farmakol. Alkaloidoi Glukozidoi* 222, (1969) *Chem. Abstr.* **70**, 22304 u

¹⁷⁴ SOKOLOV, S. YA. (1965) *Byul. Eksperim. Biol. Med.* **60**, 73, (1966) *Chem. Abstr.* **64**, 3616

¹⁷⁵ LAPIK, A. S., BUKHAROV, V. G., TALAN, V. A. and KARLIN, V. V. (1968) *Farmakol. Toksikol.* **3**, 650 (1969) *Chem. Abstr.* **70**, 56100 v

¹⁷⁶ RAMASWAMY, A. S., PIRIYASAMY, S. M. and BASU, N. K. (1970) *J. Res. Indian Med.* **4**, 160

¹⁷⁷ IMAOKA, I. (1970) *Shioku Igaku Zasshi* **26**, 1, (1970) *Chem. Abstr.* **73**, 43769

¹⁷⁸ SANDERMANN, W. and FUNK, H. (1970) *Naturwiss.* **57**, 407

¹⁷⁹ TSCHISCHE, R., WULF, G., WEBER, A. and SCHMIDT, H. (1970) *Z. Naturforsch. B* **25**, 999

¹⁸⁰ RAIPHE, A. N. Brit. 1083815 (Cl. A61K) Sept. 20, 1967, Appl. Oct. 1, 1963 (1968) *Chem. Abstr.* **68**, 33197

their action on the rectal and abdominal muscles of frog have been determined.¹⁸¹ Recently two saponins have been isolated from *Gleditschia horrida* which show antifertility effect in female mice.¹⁸²

The saponins from sea-cucumbers exhibit varied types of biological activities. Holotoxin, a crystalline saponin m.p. 250°d, isolated from the body-wall tissues of *Stichopus japonicus* showed high activity against various fungi including vegetable pathogens (*in vitro* MIC = 2–16 µg/ml) but it was almost inactive against Gram negative and Gram positive bacteria and mycobacteria. The IR spectrum of holotoxin is similar to that of holothurin but it is not sulphate. Holothurin did not exhibit antifungal activity.¹⁸³ Holothurin A caused almost complete inhibition of protein synthesis in rat bone marrow cell cultures at 10 mg/ml, stichoposides A₁, A and cucumarioside C produced 90, 70 and 50% inhibition but plant saponins, panaxosides B and C, were found to be almost completely inactive at this dose level.¹⁸⁴ Holothurin B at a concentration of 2.5×10^{-5} M, produced irreversible blockade of the responses both to indirect electrical stimulation via the nerve in the rat phrenic nerve diaphragm preparation and direct electric stimulation of muscle.¹⁸⁵ The toxic effects of stichoposide A have been studied and it was found to arrest the division of eggs of sea urchin, *Strongylocentrotus intermedius* with cytolysis of blastomeres.¹⁸⁶

¹⁸¹ BELLET, P and CEREDÉ, J ROUSSEL—UCLAF Fr 1,477,421, April 21, 1967, Appl July 20, 1964; (1967) *Chem Abstr* **67**, 102775

¹⁸² CHOU, S C, RAMANATHAN, S, MATUI, A, ROGERS, J and CUTTING, W C (1971) *Indian J Exp Biol* **9**, 503

¹⁸³ SCHIMADA, S (1969) *Science* **163**, 1462.

¹⁸⁴ ANISINOV, M M, PROKOFEVA, N G, KUZNETSOVA, T A and PERETOLCHIN, N V (1971) *Izv Akad Nauk SSR, Ser, Biol* **137**, (1971) *Chem Abstr* **74**, 73825 k, ELYAKOV, G B, ANISINOV, M M, PROKOFEVA, N G, KUZNETSOVA, T A and FRONERT, E B (1972) *Toxicon* **10**, 299, (1972) *Chem Abstr* **77**, 57344

¹⁸⁵ FRIESS, S L, DURANT, R C and CHANLEY, J D (1968) *Toxicon* **6**, 21, (1969) *Chem Abstr* **69**, 104786 b

¹⁸⁶ ANISINOV, M M, KUZNETSOVA, T A, SHIROKOV, V P, PROKOFEVA, N G and ELYAKOV, G B (1972) *Toxicon* **10**, 187, (1972) *Chem Abstr* **76**, 136599